

**NECROPSY PANELS**
**RAT**

ABBREVIATION	AGENT		CORE	ROUTINE	COMPLETE	COMPLETE PLUS
			80009	80016	80010	80025
		With Histopathology	80012	80015	80013	80026
<b>SEROLOGY</b>						
KRV	<b>Kilham's Rat Virus</b>		•	•	•	•
H-1	<b>Toolan's H-1 Virus</b>		•	•	•	•
RCAR	<b>Pneumocystis carinii</b>		•	•	•	•
SDA/RCV	<b>Rat Coronavirus</b>		•	•	•	•
RMV	<b>Rat Minute Virus</b>		•	•	•	•
RPV	<b>Rat Parvovirus</b>		•	•	•	•
RTV	<b>Rat Theilovirus</b>		•	•	•	•
CARB	<b>Cilia Associated Respiratory Bacillus</b>			•	•	•
LCMV	<b>Lymphocytic Choriomeningitis Virus</b>			•	•	•
MAV-1 (FL)	<b>Mouse Adenovirus</b>			•	•	•
MPUL	<b>Mycoplasma pulmonis</b>			•	•	•
PVM	<b>Pneumonia Virus of Mice</b>				•	•
REO	<b>Respiratory Enteric Virus III</b>				•	•
SEV	<b>Sendai Virus</b>				•	•
CPILI	<b>Clostridium piliforme</b>					•
ECUN	<b>Encephalitozoon cuniculi</b>					•
HANT	<b>Hantaan Virus</b>					•
IDIR	<b>Rat Rotavirus</b>					•
<b>PCR</b>						
HELICO	<b>Helicobacter spp.</b>		•	•	•	•
<b>MICROBIOLOGY</b>						
BBRO	<b>Bordetella bronchiseptica</b>		•	•	•	•
CKUT	<b>Corynebacterium kutscheri</b>		•	•	•	•
KOXY	<b>Klebsiella oxytoca</b>		•	•	•	•
KPNE	<b>Klebsiella pneumoniae</b>		•	•	•	•
PMUL	<b>Pasteurella multocida</b>		•	•	•	•
RPNEU	<b>Rodentibacter pneumotropica</b>		•	•	•	•
RHEYL	<b>Rodentibacter heylII</b>		•	•	•	•
PAER	<b>Pseudomonas aeruginosa</b>		•	•	•	•
SAUR	<b>Staphylococcus aureus</b>		•	•	•	•
SBETA	<b>Streptococcus spp. beta-hemolytic</b>		•	•	•	•
SPNE	<b>Streptococcus pneumoniae</b>		•	•	•	•
CROD	<b>Citrobacter rodentium</b>		•	•	•	•
SALM	<b>Salmonella spp.</b>		•	•	•	•
PMIR	<b>Proteus mirabilis</b>		•	•	•	•
CBOV	<b>Corynebacterium bovis</b>		•	•	•	•
	<b>Other Bacteria</b>		•	•	•	•

**NECROPSY PANELS  
RAT (CONT'D)**

ABBREVIATION	AGENT		CORE	ROUTINE	COMPLETE	COMPLETE PLUS
		With Histopathology	80009	80016	80010	80025
		Without Histopathology	80012	80015	80013	80026
<b>PARASITOLOGY</b>						
	<b>Ectoparasite Exam</b>		•	•	•	•
	<b>Endoparasite Exam</b>		•	•	•	•
<b>HISTOPATHOLOGY</b>						
	<b>Lung</b>		•	•	•	•
	<b>Liver</b>		•	•	•	•
	<b>Colon</b>		•	•	•	•
	<b>Cecum</b>		•	•	•	•
	<b>Harderian Gland</b>		•	•	•	•
	<b>Salivary Glands</b>		•	•	•	•
	<b>Kidney</b>		•	•	•	•
	<b>Lesioned Organs</b>		•	•	•	•

**INSTRUCTIONS FOR SUBMISSION OF SAMPLES FOR NECROPSY PANEL TESTING (IF ANIMAL NOT SENT LIVE)**

1. Euthanize the animal(s) according to your facility's IACUC guidelines.
2. Draw whole blood and spin down in a centrifuge tube to separate and then collect the serum. Alternatively, a SeraSorb™ micro-sampling device can be used as a dried whole blood collection method for serological testing. The blood draw can be taken prior to euthanasia, or immediately afterward.  
NOTE: If serology work is being requested, the blood draw MUST be taken either prior to euthanasia, or immediately afterward. If the euthanized animal is sent without blood being drawn immediately, the blood will be coagulated and therefore useless for serology by the time it arrives at VRL.
3. Additional samples can be taken prior to necropsy, depending on services required:
  - a. Bacteriological services: Take several fecal pellets and an oral swab.
  - b. Parasitology services: Take several fecal pellets and use tape to perform a fur pluck.
  - c. PCR services: Take fecal pellets and use tape to perform a fur pluck.
4. If histopathology services are required, perform an immediate necropsy. Open the abdominal cavity and the chest cavity, make observations and notes. Collect the organs in a methodical manner, usually starting with lungs, heart, liver, intestines, kidney, pancreas, lymph nodes, etc. These organs are often large, so you can trim them at necropsy with a sharp bladed instrument down to a size that will fit into a normal cassette. If using cassettes, label the cassettes with a permanent marker. Place organs/tissues (either in cassettes or not) in a jar of 10% neutral buffered formalin at 10 to 20 times the amount of tissue. Use the formalin liberally.
5. Allow the tissues to incubate in formalin for 48 - 72 hours, if being sent without trimming. If samples are trimmed thin enough to fit into a cassette, incubation for 24 - 48 hours is sufficient.
6. To ship:
  - a. For either untrimmed organs/tissues or samples in cassettes, place the jar(s) in moisture-proof containers, such as a ziploc bag, and ship to VRL.
  - b. If sending cassettes, as an alternative method to save on shipping costs, the cassettes can be placed in formalin-soaked gauze that will remain moist during shipment. Pour off excess formalin, until the gauze is wet and can keep the tissues wet. Place them in a moisture proof container and ship to VRL.