

NECROPSY PANELS

Comprehensive Animal Health Assessment Profiles

Live animal submissions (required for mouse, rat, hamster): Includes an initial clinical exam of the animal, gross necropsy, oropharyngeal and fecal bacterial culture for pathogens, pelt exam for ectoparasites, examination of the feces for helminths and microscopic examination of the duodenum, ileum, cecum and colon contents for protozoa.

Rabbits: VRL cannot euthanize rabbits. To prepare rabbit samples for necropsy panels, [follow instructions on page 63](#).

Please contact your representative if customization is needed.

ANIMAL HEALTH MONITORING

MOUSE

ABBREVIATION	AGENT	With Histopathology	CORE	ROUTINE	COMPLETE	COMPLETE PLUS
		Without Histopathology	80001	80002	80003	80004
			80005	80006	80007	80008
SEROLOGY						
MHV	Mouse Hepatitis Virus	•	•	•	•	•
MVM (MMV)	Mouse Minute Virus	•	•	•	•	•
MPV (1-5)	Mouse Parvovirus	•	•	•	•	•
MNV	Murine Norovirus	•	•	•	•	•
EDIM	Rotavirus/Epizootic Diarrhea of Infant Mice	•	•	•	•	•
TMEV/GDVII	Theiler's Mouse Encephalomyelitis Virus	•	•	•	•	•
LCMV	Lymphocytic Choriomeningitis Virus			•	•	•
MAV-1 (FL)	Mouse Adenovirus-1 (MAV-FL)			•	•	•
MAV-2 (K87)	Mouse Adenovirus-2 (MAV-K87)			•	•	•
PVM	Pneumonia Virus of Mice			•	•	•
SEV	Sendai Virus			•	•	•
CARB	Cilia Associated Respiratory Bacillus				•	•
CPILI	Clostridium piliforme				•	•
ECTRO	Ectromelia Virus				•	•
MCMV	Murine Cytomegalovirus				•	•
MYCO	Mycoplasma pulmonis				•	•
POLY	Polyomavirus				•	•
REO	Respiratory Enteric Virus III				•	•
ECUN	Encephalitozoon cuniculi					•
HANT	Hantaan Virus					•
KV	K Virus (Mouse Pneumonitis Virus)					•
LDHV	Lactate Dehydrogenase Elevating Virus					•
MTV	Mouse Thymic Virus					•
PCR						
HELICO	Helicobacter spp.		•	•	•	•
HISTOPATHOLOGY						
	Lung		•	•	•	•
	Liver		•	•	•	•
	Colon		•	•	•	•
	Cecum		•	•	•	•
	Lesioned Organs		•	•	•	•

**NECROPSY PANELS
MOUSE (CONT'D)**

ABBREVIATION	AGENT		CORE	ROUTINE	COMPLETE	COMPLETE PLUS
			80001	80002	80003	80004
		With Histopathology	80001	80002	80003	80004
		Without Histopathology	80005	80006	80007	80008
PARASITOLOGY						
	Ectoparasite Exam		•	•	•	•
	Endoparasite Exam		•	•	•	•
MICROBIOLOGY						
BBRO	Bordetella bronchiseptica		•	•	•	•
CKUT	Corynebacterium kutscheri		•	•	•	•
KOXY	Klebsiella oxytoca		•	•	•	•
KPNE	Klebsiella pneumoniae		•	•	•	•
PMUL	Pasteurella multocida		•	•	•	•
RPNEU	Rodentibacter pneumotropica		•	•	•	•
RHEYL	Rodentibacter heylii		•	•	•	•
PAER	Pseudomonas aeruginosa		•	•	•	•
SAUR	Staphylococcus aureus		•	•	•	•
SBETA	Streptococcus spp. beta-hemolytic		•	•	•	•
SPNE	Streptococcus pneumoniae		•	•	•	•
CROD	Citrobacter rodentium		•	•	•	•
SALM	Salmonella spp.		•	•	•	•
PMIR	Proteus mirabilis		•	•	•	•
CBOV	Corynebacterium bovis		•	•	•	•
	Other Bacteria		•	•	•	•

INSTRUCTIONS FOR SUBMISSION OF SAMPLES FOR NECROPSY PANEL TESTING (IF ANIMAL NOT SENT LIVE)

1. Euthanize the animal(s) according to your facility's IACUC guidelines.
2. Draw whole blood and spin down in a centrifuge tube to separate and then collect the serum. Alternatively, a SeraSorb™ micro-sampling device can be used as a dried whole blood collection method for serological testing. The blood draw can be taken prior to euthanasia, or immediately afterward.
NOTE: If serology work is being requested, the blood draw MUST be taken either prior to euthanasia, or immediately afterward. If the euthanized animal is sent without blood being drawn immediately, the blood will be coagulated and therefore useless for serology by the time it arrives at VRL.
3. Additional samples can be taken prior to necropsy, depending on services required:
 - a. Bacteriological services: Take several fecal pellets and an oral swab.
 - b. Parasitology services: Take several fecal pellets and use tape to perform a fur pluck.
 - c. PCR services: Take fecal pellets and use tape to perform a fur pluck.
4. If histopathology services are required, perform an immediate necropsy. Open the abdominal cavity and the chest cavity, make observations and notes. Collect the organs in a methodical manner, usually starting with lungs, heart, liver, intestines, kidney, pancreas, lymph nodes, etc. These organs are often large, so you can trim them at necropsy with a sharp bladed instrument down to a size that will fit into a normal cassette. If using cassettes, label the cassettes with a permanent marker. Place organs/tissues (either in cassettes or not) in a jar of 10% neutral buffered formalin at 10 to 20 times the amount of tissue. Use the formalin liberally.
5. Allow the tissues to incubate in formalin for 48 - 72 hours, if being sent without trimming. If samples are trimmed thin enough to fit into a cassette, incubation for 24 - 48 hours is sufficient.
6. To ship:
 - a. For either untrimmed organs/tissues or samples in cassettes, place the jar(s) in moisture-proof containers, such as a ziploc bag, and ship to VRL.
 - b. If sending cassettes, as an alternative method to save on shipping costs, the cassettes can be placed in formalin-soaked gauze that will remain moist during shipment. Pour off excess formalin, until the gauze is wet and can keep the tissues wet. Place them in a moisture proof container and ship to VRL.