

**NECROPSY PANELS**
**HAMSTER**

ABBREVIATION	AGENT	ROUTINE COMPLETE	
		With Histopathology 80030	80033
		Without Histopathology 80027	80034
<b>SEROLOGY</b>			
LCMV	<b>Lymphocytic Choriomeningitis Virus</b>	•	•
PVM	<b>Pneumonia Virus of Mice</b>	•	•
REO	<b>Respiratory Enteric Virus III</b>	•	•
SEV	<b>Sendai Virus</b>	•	•
SV5	<b>Simian Virus 5</b>	•	•
CPILI	<b>Clostridium piliforme</b>		•
ECUM	<b>Encephalitozoon cuniculi</b>		•
<b>MICROBIOLOGY</b>			
BBRO	<b>Bordetella bronchiseptica</b>	•	•
CKUT	<b>Corynebacterium kutscheri</b>	•	•
KOXY	<b>Klebsiella oxytoca</b>	•	•
KPNE	<b>Klebsiella pneumoniae</b>	•	•
PMUL	<b>Pasteurella multocida</b>	•	•
RPNEU	<b>Rodentibacter pneumotropica</b>	•	•
RHEYL	<b>Rodentibacter heylii</b>	•	•
PAER	<b>Pseudomonas aeruginosa</b>	•	•
SAUR	<b>Staphylococcus aureus</b>	•	•
SBETA	<b>Streptococcus spp. beta-hemolytic</b>	•	•
SPNE	<b>Streptococcus pneumoniae</b>	•	•
CROD	<b>Citrobacter rodentium</b>	•	•
SALM	<b>Salmonella spp.</b>	•	•
PMIR	<b>Proteus mirabilis</b>	•	•
CBOV	<b>Corynebacterium bovis</b>	•	•
	<b>Other Bacteria</b>	•	•
<b>PARASITOLOGY</b>			
	<b>Ectoparasite Exam</b>	•	•
	<b>Endoparasite Exam</b>	•	•
<b>HISTOPATHOLOGY</b>			
	<b>Lung</b>	•	•
	<b>Liver</b>	•	•
	<b>Colon</b>	•	•
	<b>Cecum</b>	•	•
	<b>Kidney</b>	•	•
	<b>Lesioned Organs</b>	•	•

**INSTRUCTIONS FOR SUBMISSION OF SAMPLES FOR NECROPSY PANEL TESTING (IF ANIMAL NOT SENT LIVE)**

1. Euthanize the animal(s) according to your facility's IACUC guidelines.
2. Draw whole blood and spin down in a centrifuge tube to separate and then collect the serum. Alternatively, a SeraSorb™ micro-sampling device can be used as a dried whole blood collection method for serological testing. The blood draw can be taken prior to euthanasia, or immediately afterward.  
NOTE: If serology work is being requested, the blood draw MUST be taken either prior to euthanasia, or immediately afterward. If the euthanized animal is sent without blood being drawn immediately, the blood will be coagulated and therefore useless for serology by the time it arrives at VRL.
3. Additional samples can be taken prior to necropsy, depending on services required:
  - a. Bacteriological services: Take several fecal pellets and an oral swab.
  - b. Parasitology services: Take several fecal pellets and use tape to perform a fur pluck.
  - c. PCR services: Take fecal pellets and use tape to perform a fur pluck.
4. If histopathology services are required, perform an immediate necropsy. Open the abdominal cavity and the chest cavity, make observations and notes. Collect the organs in a methodical manner, usually starting with lungs, heart, liver, intestines, kidney, pancreas, lymph nodes, etc. These organs are often large, so you can trim them at necropsy with a sharp bladed instrument down to a size that will fit into a normal cassette. If using cassettes, label the cassettes with a permanent marker. Place organs/tissues (either in cassettes or not) in a jar of 10% neutral buffered formalin at 10 to 20 times the amount of tissue. Use the formalin liberally.
5. Allow the tissues to incubate in formalin for 48 - 72 hours, if being sent without trimming. If samples are trimmed thin enough to fit into a cassette, incubation for 24 - 48 hours is sufficient.
6. To ship:
  - a. For either untrimmed organs/tissues or samples in cassettes, place the jar(s) in moisture-proof containers, such as a ziploc bag, and ship to VRL.
  - b. If sending cassettes, as an alternative method to save on shipping costs, the cassettes can be placed in formalin-soaked gauze that will remain moist during shipment. Pour off excess formalin, until the gauze is wet and can keep the tissues wet. Place them in a moisture proof container and ship to VRL.